



คณะแพทยศาสตร์ศิริราชพยาบาล มหาวิทยาลัยมหิดล

## Diagnosis for infectious diseases

### “Molecular based techniques”

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## Real-life situations

- Hemoculture grows bacterial colonies on blood agar
- Automated blood culture flagged positive
- CSF sample from patients with meningoencephalitis
  - Culture: no growth or previous antibiotic Rx
- Acid fast positive from clinical specimens or from mycobacterial culture

### How molecular techniques assist our practice?

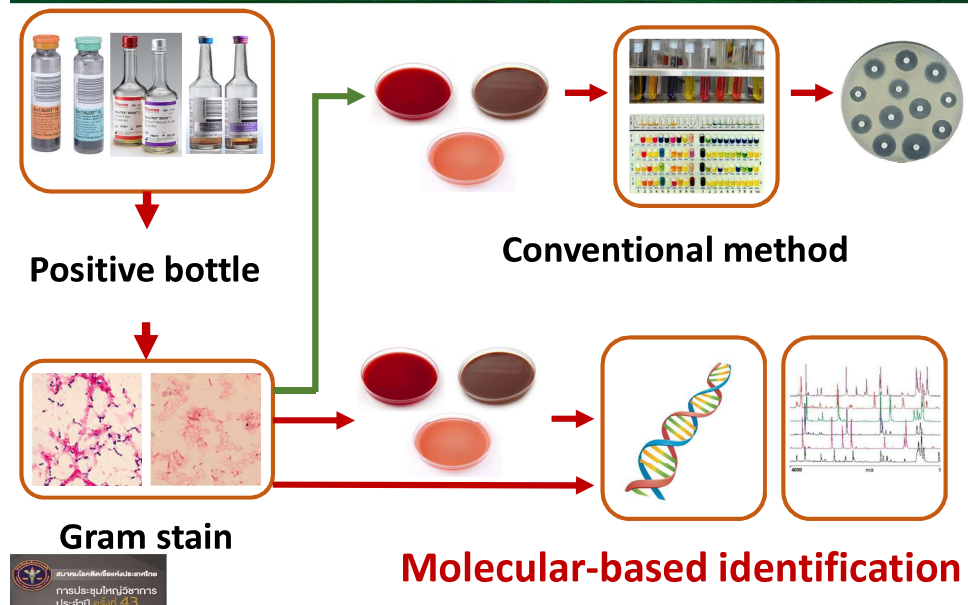


## Outline

- Culture-based molecular identification
- Direct molecular detection

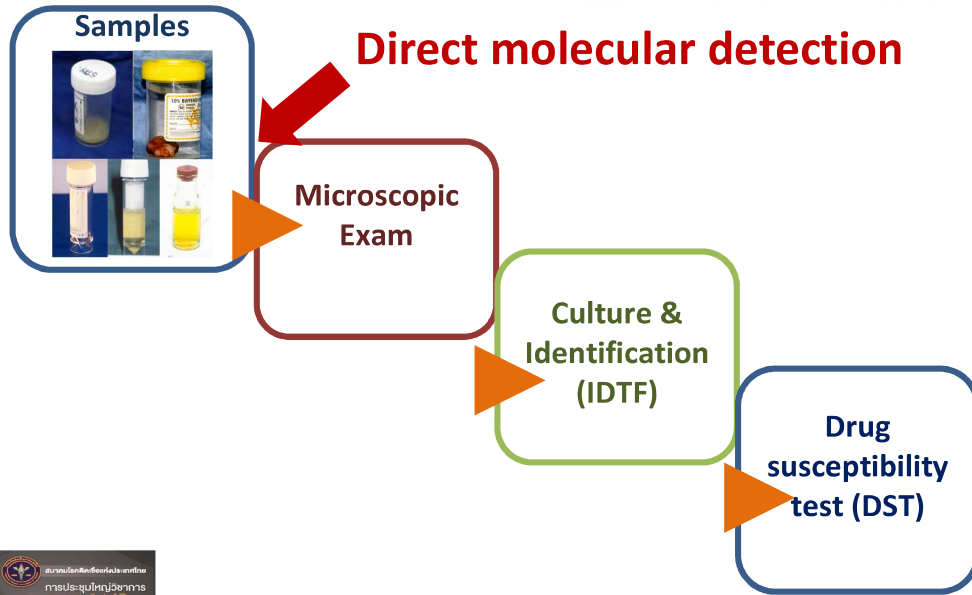


## Culture-based molecular identification

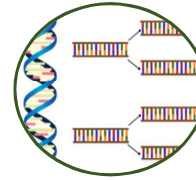




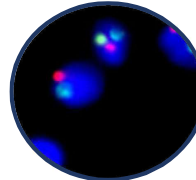
## Direct molecular detection



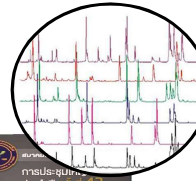
## Basic molecular techniques for organism identification



**Amplified nucleic acid-based**  
e.g. polymerase chain reaction (PCR)



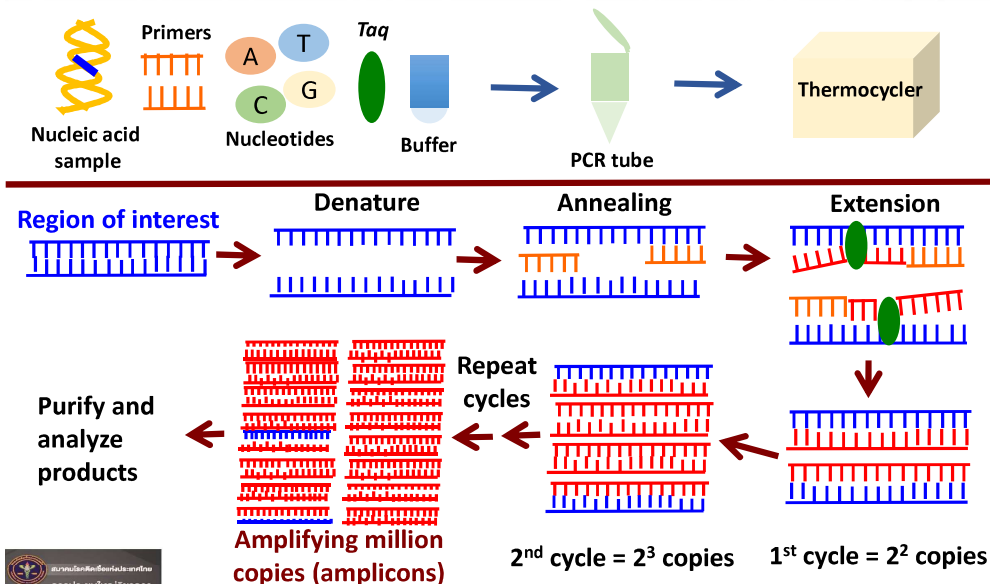
**Nucleic acid hybridization**  
e.g. fluorescent in situ hybridization (FISH),  
line probe assay, microarray



**Non-nucleic acid-based**  
e.g. Mass spectrometry

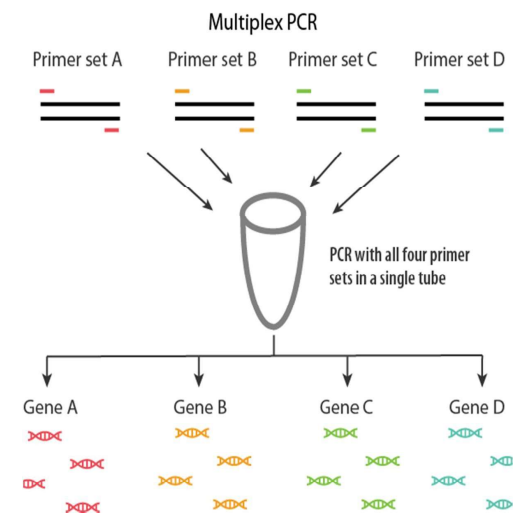


## PCR-based method: Basic concept



## Multiplex PCR

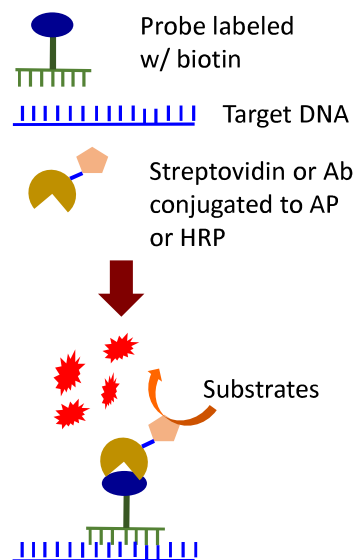
- Use > 1 primer sets at once
- Target multiple organisms or genes
- Fluorescence discriminates each amplicon product
- Useful application in clinical microbiology
  - Respiratory sample, CSF, stool, genital swab





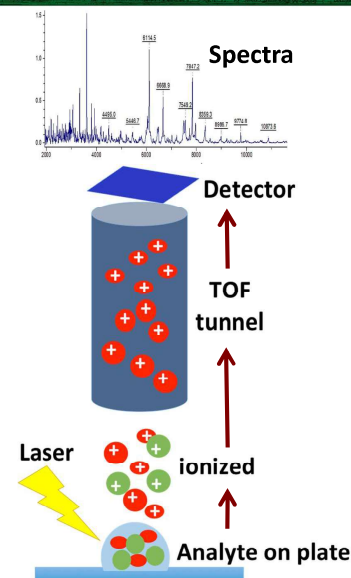
## Nucleic-acid hybridization

- Use probes to identify target DNA or RNA and are labelled with biotin or digoxigenin
- Probes hybridize to the target
- Detect the hybridization by conjugated enzyme/substrate reactions
  - Color change (chromogenic)
  - Chemiluminescent or light signals under optical camera



## Non-nucleic acid-based: Matrix-assisted laser desorption ionization time-of-flight mass spectrometry (MALDI-TOF MS)

- Analyte preparation (sample + matrix)
- Laser desorb the analyte on plate into ionized molecules, and run through an electrostatic tunnel
- Small molecules go fast, followed by larger molecules (time-of flight)
- The molecules detected by an ion detector
- Mass spectra are unique to specific genera and species



Clin Microbiol Infect 2010;16:1604-13., Clin Infect Dis 2013;57:564-72.



## MALDI-TOF MS

- 2 commercial MALDI-TOF systems

**Biotyper (Bruker, USA)**



**Vitek MS (Biomerieux, France)**



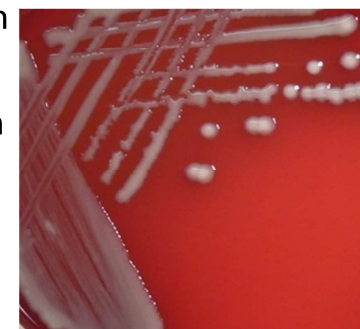
- Overall performance is comparable
- Differences are associated with software databases

Future Microbiol 2014;9:543-59.



## Case 1

- A patient has a prolonged admission in an ICU
- Previously receiving broad spectrum antibiotics for weeks
- Previous colonization with XDR *A. baumannii* (sputum) and CRE *K. pneumoniae* (urine)
- The patient has a new onset of sepsis
- Blood culture grew the isolate as shown



**Blood agar plate**

**Any methods for early organism identification?**



## Case 2

- A patient, Dx AML post CMT, has prolonged neutropenia, fever and mucositis
- On meropenem, colistin, vancomycin and amphotericin B
- New onset of sepsis with shock
- Automated blood Cx is alarmed at 4 hrs; Gram stain as shown



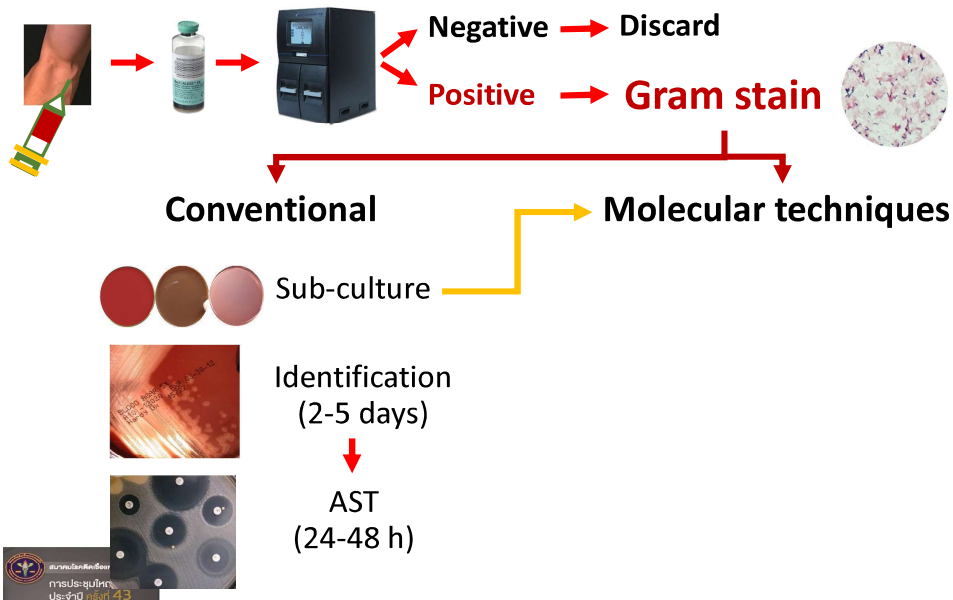
Gram stain from blood culture bottle

**Any methods for early organism identification?**

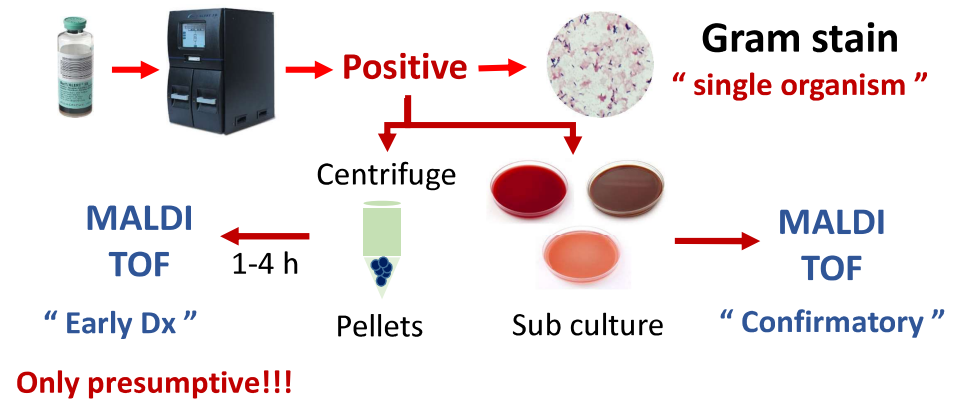


## Exploitation of molecular techniques in blood culture positive

### Identifying organisms from positive blood culture



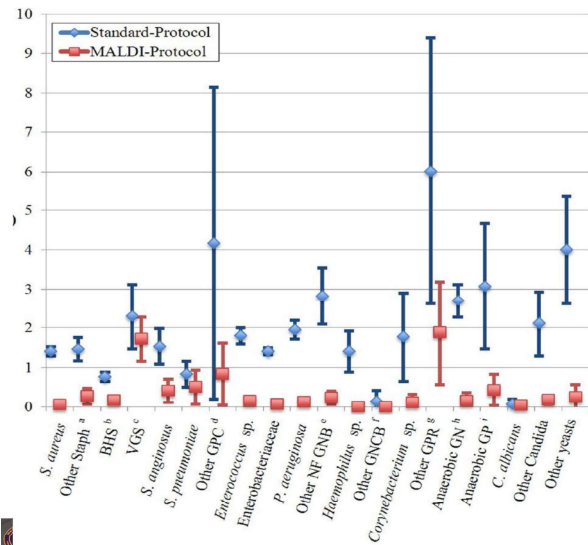
### MALDI-TOF MS for positive blood culture





## Time to identification (TTI) and cost effectiveness studies

TTI (Days)



The MALDI provided the identifications

- 1.45 days earlier ( $P < 0.001$ )

Cost effectiveness

Cost/12 months	Standard protocol (\$)	MALDI (\$)
Reagents	158,645	29,613
Labor	31,323	26,669
Fixed annual		31,272
<b>Total</b>	<b>189,969</b>	<b>87,555</b>

Save cost 53.9%

Tan Tien et al. Clin Microbiol 2012;50:3301-8.



## MALDI-TOF MS for positive blood culture

### Limitations

- Polymicrobial infection
- Anaerobic bacteria
- GNR non fermenters
- Closely related species such as
  - *Streptococcus pneumoniae* vs. *S. mitis* group
  - *Bacillus* spp.
  - *E. coli* vs. *Shigella*
  - *Candida* spp.

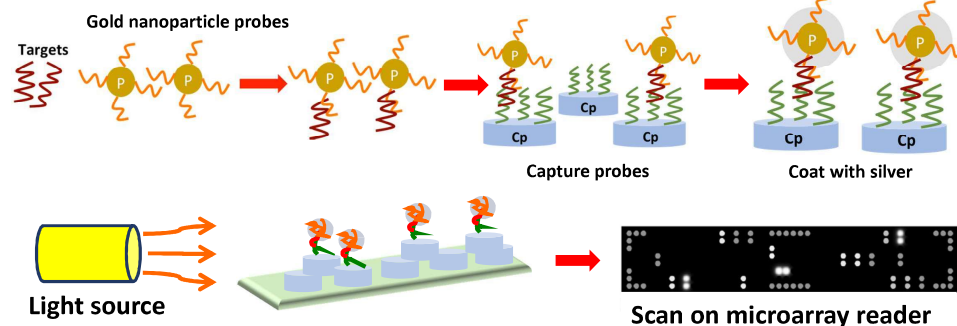
**"Only preliminary report, require back-up from conventional identification"**

Clin Microbiol Rev 2013;26:547-603.  
Future Microbiol 2014;9:543-59., Clin Microbiol Infect 2015;21:313-22.

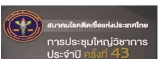


## Microarray-based for positive blood culture

- Probes and targets hybridization on microarray glass slide
- Detect the targets by an optical camera in the microarray reader



Clin Microbiol Rev 2014;27:783-822., Clin Chem Lab Med 2015;53:1013-24.



## Commercial microarray for positive blood culture

- Reduce the cost per target tested
- Simultaneous testing for multiple pathogens with similar symptoms

Test	Targets	Sensitivity (%)	Specificity (%)	Time to result (h)
<b>Verigene BC-GP</b>	12 Gram positive, 3 resistance genes ( <i>mecA</i> , <i>vanA</i> , <i>vanB</i> )	92-100	98-100	2.5
<b>Verigene BC-GN</b>	8 Gram negative, 6 resistance genes (KPC, NDM, CTX-M, VIM, IMP, OXA)	81-100	98-100	2
<b>Prove-it Sepsis</b>	60 bacteria, 13 fungi, <i>mecA</i>	95%	99%	3.5

### Limitations

- *S. pneumoniae*, polymicrobial infection

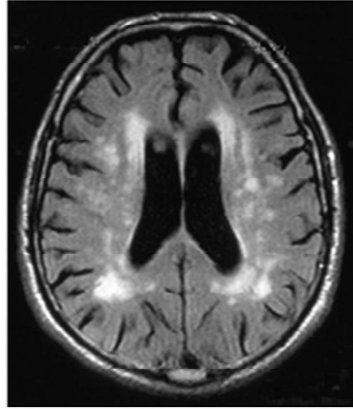
Clin Microbiol Rev 2014;27:783-822., Clin Microbiol Infect 2015;21:313-22.



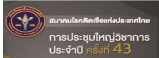


## Case 3

- A patient with SLE with active LN receiving prednisolone and cyclophosphamide Rx
- She developed altered mental status and seizure
- CSF culture: No growth
- MRI brain suspected infectious meningoencephalitis vs. vasculitis



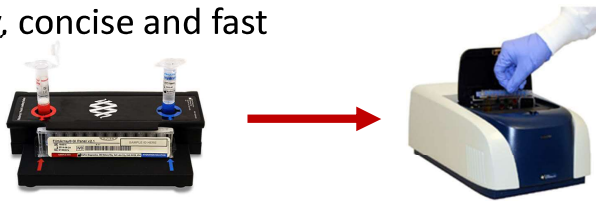
**Any molecular methods for identifying infectious etiology?**



## Exploitation of molecular techniques in CNS infection

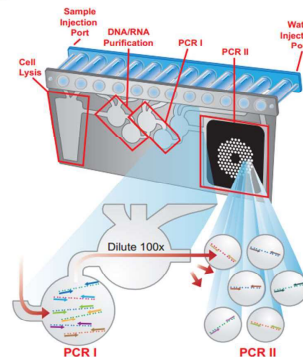
### Automated multiplex real time PCR (BioFire FilmArray®)

- Easy, concise and fast



- A qualitative method using 2 stages PCR

- 1<sup>st</sup> PCR to enrich the target
- 2<sup>nd</sup> PCR using specific primers, detect the target by fluorescent dye



<http://www.biofire.com/wp-content/uploads/2016>

### BioFire FilmArray® Meningitis/Encephalitis panel

- Uncentrifuged CSF specimens at least 200 µL
- Identify the following organisms

*S. pneumoniae*  
*S. agalactiae*  
*N. meningitides*  
*L. monocytogenes*  
*E. coli* K1  
*H. influenzae*

CMV  
 Enterovirus  
 HSV-1  
 HSV-2  
 HHV-6  
 VZV  
 Human parechovirus

*C. neoformans/gattii*

#### • Limitations

- Latent vs. active CMV and HHV-6 infections
- Non-K1 *E. coli*, non encapsulated *N. meningitides*
- CSF samples from shunt devices
- Data in immunocompromised populations



<http://www.biofire.com/wp-content/uploads/2016>



## Multicenter study of BioFire FilmArray ME Panel

- **1,560 CSF specimens**
  - Positive 136 (8.7%), negative 1,424 (91.3%),
  - 84.4% positive and >99.9% negative agreement with standard
- **False POSITIVE ME panel (22)**
  - *S. pneumoniae* (7), HHV-6 (3), CMV (2), EBV (2), HSV-1 (2), *C. neoformans/gattii* (2)
  - *E. coli* (1), *S. agalactiae* (1), HSV-2 (1), VZV (1)

**“CSF culture or conventional methods are still needed, regardless of the FilmArray ME Panel”**

*J Clin Microbiol* 2016;54:2251-61.

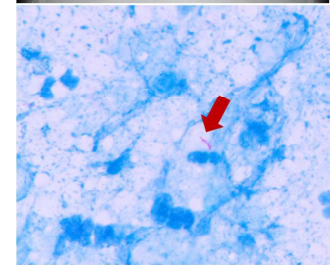


## Case 4

- A 16-year-old woman has pneumonia and ARDS
- Her symptoms was not responsive after IV cefepime and azithromycin Rx
- Sputum culture: no growth
- Sputum AF stain as shown

**Is it likely MTB?**

**Any molecular methods for Dx MTB?**



## Direct molecular detection

### New molecular detection

- Detection of mycobacterial DNAs
- Amplify the targets by PCR assay
- Presumptive diagnosis

### Xpert MTB/RIF®

- Approved by WHO since 2010
- Semi-automated technique

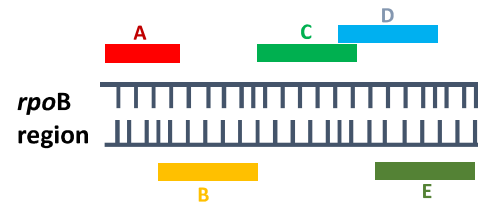
### Samples



Culture and identification  
(2-6 weeks)



## Xpert MTB/RIF



- Target: MTB *rpoB* gene
- PCR and detect the target by 5 specific probes (A-E)
- Semi-quantitative assay
- Detect *rpoB* gene mutation

### Semi-automated technique



**Turn-around time ≈ 2 hrs**

WHO. Policy update, 2013.





## Xpert MTB/RIF

### For Dx pulmonary TB in adults

	Pooled sensitivity	Pooled specificity
Initial test regardless of smear	88%	99%
Add-on test following smear (-)	68%	99%
Detect RIF resistance	95%	98%

### For Dx extra-pulmonary TB

	Pooled sensitivity	Pooled specificity
Pleural fluid	17-43%	98-99%
CSF	55-79%	98%
LN Bx and aspirate	83-85%	92-99%
Tissue BX	81%	98%

WHO. Policy update, 2013.



## Limitations

- Presumptive diagnosis
- Pediatric and other samples has not been validated
- A positive test does not indicate viable organisms
- False negativity (low MTB load, post treatment, or processing errors)
- False positivity (*M. scrofulaceum*,  $\geq 10^8$  CFU/mL)



Xpert® MTB/RIF. Package insert, Cepheid 2015.



## Case 5

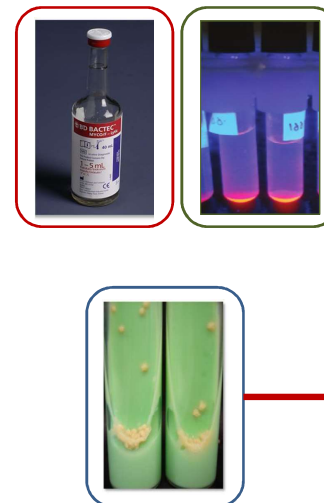
- A bottle from an automated mycobacterial liquid media culture alarmed positive
- Gram stain and AF stain compatible with mycobacteria

Is it MTB or NTM?

Any molecular methods for mycobacteria species identification?



## Hybridization technique for mycobacterial identification



### Molecular identification assays

- Gold standard
- Provide early diagnosis

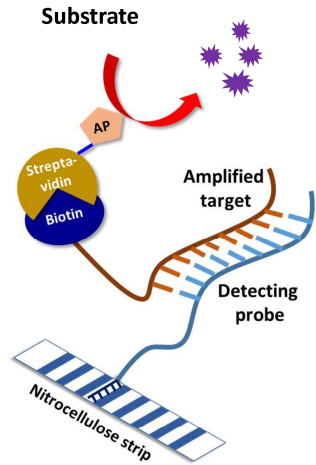
### Line probe assays

- Amplify targets by PCR
- Pretreat the amplicons with biotin
- Probe hybridize with target DNA or RNA
- Detect color change or fluorescence
- Commercial kits
  - INNO-LiPA®, GenoType®

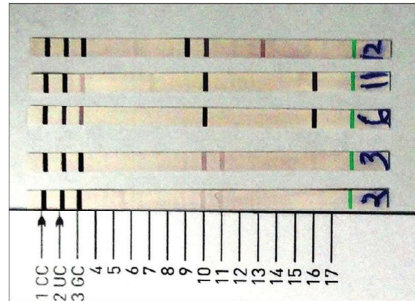




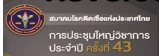
## Line probe assay



- Species specific probes immobilized on nitrocellulose strip
- Biotinylated amplicons hybridize with the probes
- Streptavidin-AP reacts with substrate  
→ purple brown band



“Reverse hybridization”

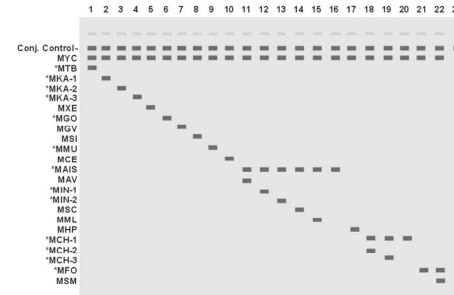


INNO-LiPA MYCOBACTERIA v2, Innogenetics, *Journal of Laboratory Physicians*, 2013;5:83-9



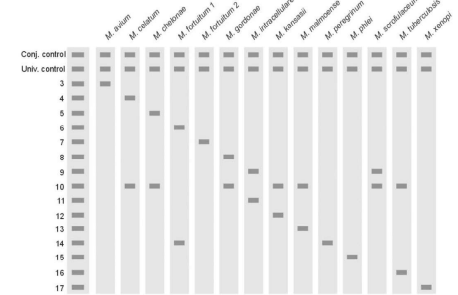
## INNO-LiPA® vs. GenoType®

### INNO-LiPA®



Sensitivity 100%, specificity 94.4%

### GenoType® CM



Sensitivity 97.9%, specificity 92.4%

Uncommon mycobacteria can be misidentified as *M. fortuitum* or *M. avium-intracellulare*

None of the systems misidentified MTB complex



Tortoli E, et al. *J Clin Microbiol* 2003;41:4418-20., Russo C, et al. *J Clin Microbiol* 2006;44:334-9., Tortoli E, et al. *J Clin Microbiol* 2010;48:307-10.



## Pros and Cons

### Pros

- Rapid identification
- No influence of antibiotics
- High sensitivity
- Semi-automated or automated system
- Less labor-intensive
- Appropriate for highly fastidious microorganisms

### Cons

- Lack of a gold standard
- No susceptibility data
- Detect nucleic acids instead of viable organisms
- Restricted availability
- Presence of contamination
- Cost-effectiveness?



*Lancet Infect Dis* 2004;4:751-60.  
*Clin Microbiol Infect* 2015;21:313-22.



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## Diagnosis for infectious diseases

# “ Molecular based techniques ”

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